# Identification and characterization of two new recessive embryonic lethal mutations in Holstein cattle

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1000 bull genomes project



### Introduction

Breeds are genetically small populations

Average inbreeding rate ~ 1% /generation

Regular emergence of recessive defects

Observatories + homozygosity mapping





In front of a typical pedigree ...





Efficient tools to detect novel defects and map associated mutations in a short period of time with a limited number of cases



CVM (A. Gentile)



#### Introduction



However, this process rely on the observation of clinical cases affected animals with distinctive symptoms

#### Two approaches

 $\rightarrow$  Identification of 2 lethal embryonic mutations

#### Depletion in homozygous genotypes

(VanRaden *et al.*, 2011)



Look for haplotypes with a lack in homozygous animals

Search for causative mutations using WGS

#### **Reverse genetics**

(Charlier et al., 2016)

Identify deleterious mutations heterozygous in sires' WGS and predict their effects

Add them on SNP bead chip



~ 1000 markers 120 000 animals genotyped per year

Clinical examination of homozygous









#### Material



- Previous study in 2013 by Fritz et al. on 48 000 Holstein
- ~ 150 000 Holstein genotyped (with sire and MGS <u>or</u> sire and dam genotyped)
- Process of genomic evaluation
- (1/3 50K, 2/3 EuroG10k Illumina Beadchips and imputation)

Sliding windows of 20-markers



Comparison (Chi-2) between numbers of observed and expected homozygotes

Consideration of haplotypes with : Nobs/Nexp <0.25 & adjusted p-value <0.01





#### Results

Name	ВТА	Interval (UMD3.1 Mb)	Haplotype freq. (%)	Nexp	Nobs	Chi² test
HH3	8	94.5-95.6	3.1%	332	3	7.4 x 10 <sup>-91</sup>
HH4	1	0.1-1.4	4.4%	301	8	5.9 x 10 <sup>-82</sup>
BY	21	20.0-21.2	2.7%	124	1	6.2 x 10 <sup>-30</sup>
HH5	9	94.8-96.4	1.9%	117	8	3.6 x 10 <sup>-28</sup>
HH1	5	63.0-65.6	1.7%	57	1	1.1 x 10 <sup>-11</sup>
HH6	16	27.8-32.0	1.1%	31	0	1.7 x 10 <sup>-4</sup>

Charlier et al. 2012; Fritz et al., 2013; McClure et al., 2014; Daetwyler et al. 2014; Adams et al., 2016; Schütz et al., 2016).

- Detection of previously identified BY, HH1, HH3, HH4 and HH5, no detection of HH2
- Observed homozygotes for haplotypes are heterozygous for the causative mutations
- ► Detection of a new region HH6 (freq 1.7%) 0 observed vs 31 homozygotes expected → embryonic lethal ?
- Most influential and ancient carrier is Mountain (BIS-MAY S-E-L MOUNTAIN ET)
- Fine-mapping using intra familial recombinations  $\rightarrow$  interval of 1.1 Mb





## Identification of candidate mutation

- Use of 186 Holstein bull's whole genome sequences from Run4 of the 1000 bull genome consortium (10 HH6C and 176 HH6F)
- Correlation between status on haplotypes and genotypes
  for the variants in the reduced interval





Candidate mutation : chr16 g.29773628A>G affects the initiator codon of the gene SDE2 Telomere Maintenance Homolog





### Identification of candidate mutation

- Function of SDE2 protein
  - Maintain genomic stability during mitosis
  - ▶ KO causes chromosomal rearrangements and losses in Fission Yeast
- Entirely conserved among eukaryotes: Mutant SDE2 protein truncated by 83 AA

Wildtype protein Mutant protein (predicted)

B. taurus (Cow) H. sapiens (Human) G. gallus (Chicken) P. sinensis (Chinese softshell turtle) D. rerio (Zebrafish) D. elegans (Fruit fly) C. interstinalis (Vase tunicate) O. sativa (Rice) S. pombe (Fission yeast)

	New start
	DIVQHGAVISLEPRLRGGKGGFGSMLRALGAQIEKTINREACRDLSGR
	DTVQHGAVYSLEPRLCGGKGGFGSMLRALGAQIEKTTNREACRDLSGR
	DVLRDGAVYSLELRLCGGKGGFGSMLRALGAQIEKTTNREACRDLSGR
)	DLLQDGVVYSLEPRLCGGKGGFGSMLRALGAQIEKTTNREACRDLSGR
·	EKLQDGHTYSIEPRLCGGKGGFGSMLRALGAQIEKTTNREACRDLSGR
	DLLQSGVVYRLEPRLCGGKGGFGSMLRALGAQIEKTTNREACRDLSGR
	-ELLHGDVHCVLRQL-GGKGGFGSMLRAIGAQIEKTTNREACRDLSGR
	DSVQNGDYLAAKFRLLGGKGGFGSMLRALGSQIEKTTNNEACRDLSGR
	SVLASSADGRFPSASALLRLRGGKGGFGSLLRGAASKAGQKKTSNFDACRDINGR
	IQLCKLEGKSTSAHLNLTLCTRVLGGKGGFGSQLRAAGGRMSKKRNEQENQDSCRDLDGN
	: *************************************

8-AA motif conserved

# Reverse genetic approach identifies HH7

#### Experience from large scale use of the EuroGenomics custom SNP chip in cattle

Using the Eurogenomics chip (Friday 1:30)

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- 1000 markers designed for deleterious mutations heterozygous in sires' genomes
  - → Detection of mutations intrabreed
  - → Focus on embryonic lethal
- ▶ A four base pair deletion in CENPU gene downstream the splice donor site (6 +/- VS 1141 +/+)

Chr27:14168128

ATTACT**TACT** ATTACT

- Specialized chromatin domain which play a key role in mitosis
- Close to the splice donor site of CENPU
  → modifications of the primary structure of the protein
- → Validation ongoing

Mice



#### Splicing donor site

Homozygotes for a knock-out allele exhibit embryonic lethality between E7.5 and E9.5, small embryo size and thickened visceral endoderm.





## Large scale genotyping

No homozygous to the mutation

On EuroGenomics beadchip since 2015

0 homozygous observed (out of 100 100 animals genotyped) Freq=0.9%

Comparison with the depletion in homozygous analysis

The haplotype (HH7) associated with this mutation shows significant depletion in homozygous without Bonferroni correction !!

Name	ВТА	Interval (UMD3.1 Mb)	Haplotype freq. (%)	Nexp	Nobs	Chi <sup>2</sup> test	Bonferronni corrected
HH7	27	13.0-14.4	1.2%	16	0	2.0 x 10 <sup>-6</sup>	18.6

Ancestral version of the haplotype Recombining predating the mutation event +/+ HH7/+ HH7/HH7 Total +/+ 20 98244 98224 0 +/DEL 1692 164 0 1856 DEL/DEL 0 0 0 0 Total 98388 1712 0 100100

► No homozygote for HH7 haplotype or candidate mutation





## Effects on fertility of HH6 and HH7

Decrease in fertility observed in mating among carriers



- Analysis of Conception and Non Return Rates suggests that embryos die before 35 days of gestation
- Estimated values were closed to the expected effect under the assumption of complete lethality in homozygous embryos





#### Discussion

Reverse genetics approach :

Study gene function

Lot of false-positives: (Charlier *et al.*)

- 15% deleterious
- 6% missense are true

Depletion in homozygous

Few false-positive

Need a huge population of genotyped animals to identify haplotypes with a frequency higher than 1%

Linkage Disequilibrium

→ Complementarity of the two approaches

# Thank you for your attention